

# Image analysis

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- RapiGrow
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### Introduction

**Phenotyping** is a key approach to understand how plants interact with environmental change as well as respond to different treatments

- Conventional plant phenotyping involves manual measurement of plant traits (e.g., area, biomass, height, plant growth rate, etc) which is slow, tedious and expensive task
- Recently, computer-assisted methods for phenotyping, and in particular imaging techniques are becoming more popular



### Plant phenotyping networks around the world

Network	Name	Web Location			
APPF	Australian Plant Phenotyping Facility	http://www.plantphenomics.org.au/			
APPN	Austrian Plant Phenotyping Network	http://www.appn.at/			
DPPN	German Plant Phenotyping Network	https://dppn.plant-phenotyping-network.de/			
EPPN	European Plant Phenotyping Network	https://www.plant-phenotyping-network.eu/			
FPPN	French Plant Phenotyping Network	https://www.phenome-fppn.fr/			
IPPN	International Plant Phenotyping Network	https://www.plant-phenotyping.org/			
LatPPN	Latin American Plant Phenotyping	https://www.frontiersin.org/articles/10.3389/			
	Network	fpls.2016.01729/full			
NaPPI	Finland National Plant Phenotyping	https://www.helsinki.fi/en/infrastructures/na			
	Infrastructure	tional-plant-phenotyping			
NPPN	Nordic Plant Phenotyping Network	https://www.forageselect.com/nppn			
Phen-Italy	Italian Plant Phenotyping Network	http://www.phen-italy.it/			
UKPPN	UK Plant Phenotyping Network	http://www.ukppn.org.uk/ Ashok etal			
PheNo	Norwegian Plant Phenotyping Platform	https://emphasis.plant- phenotyping.eu/News? news=Norway_Application_PheNo			



### RapiGrow

**RapiGrow** project aims to develop machine learning/deep learning models that can predict plant growth from images, and observe plant nutrient availability (e.g., phosphorus) by applying hyperspectral image analysis of plant growth

 As a case study, machine learning models will be developed to predict plant biomass from image-based features/traits (e.g., area)



### RapiGrow

The implementation approach involves the following major steps:

- Experimental setup and plant image acquisition
- Pre-processing
- Segmentation
- Feature extraction
- Prediction model development (train, validate and test)
- Prediction model evaluation



### Imaging techniques and experimental setup

- **Digital color (RGB) imaging**: structural plant traits like leaf area, stem length, plant growth rate
- **Hyperspectral imaging**: physiological plant traits like leaf and canopy water content, quantifying chemical properties

Car	meras		Hyperspectral					
	VIS	IR	NIR/SWIR			TIR/LWIR		
4	00 500 600	700 800 900 100	00 1100 1200 1300 1400 1500 1600 1700 Wavelength (nm)	1800 1900 2000 2100 220	0 2300 2400 2500	)7.5–13x10 <sup>3</sup>		
UV	Visible	Near-infrared	Short-wave	e infrared		Long-wave infrared		
		Water content	• •	-	Temperature			
		Chlorophyll	content, photosystem II quantum efficiency					
				Fahlaren etal	Current Opinion		niversity of Life Sci	ences



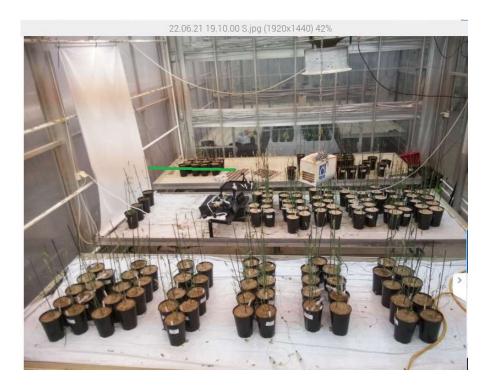
#### Imaging techniques and experimental setup

- Raspberry Pi computer and Camera module
- Automated data collection using Python



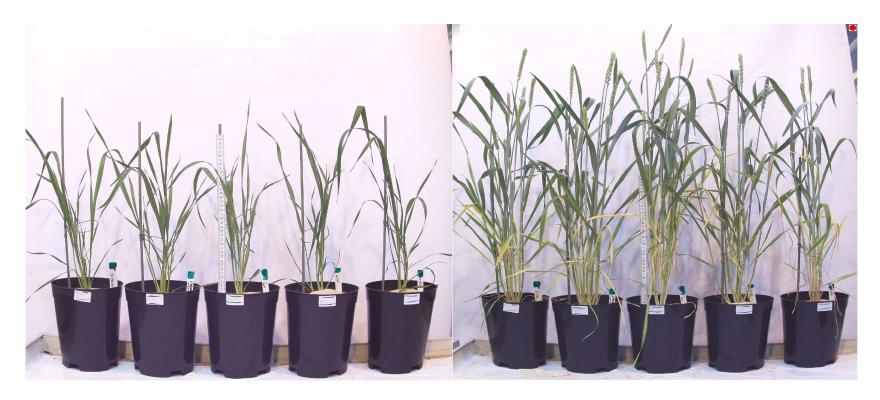


### Imaging techniques and experimental setup





### Sample images





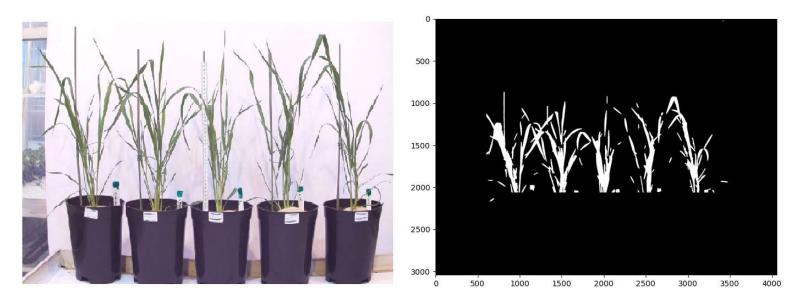
### Sample images





#### Image pre-processing and segmentation

• Different segmentation techniques (color index, threshold, and learning-based segmentation)



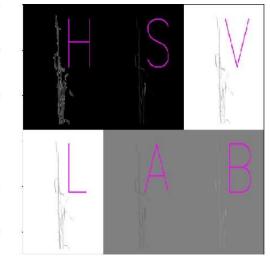


#### Image preprocessing and segmentation

Different segmentation techniques (color, Otsu's thresholding, and learning-based

segmentation)

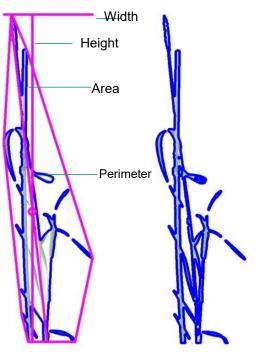






#### **Feature extraction**

• Measurement of morphological traits, such as area, perimeter, height, width, etc



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### **Feature extraction**

le Edit	Vie	w Inse	ert Cell	Kerne	al VV	idgets Help	
+ 8	2	B 🛧	↓ ► Run		C #	Code	<ul> <li>Image: Second sec</li></ul>
Out[22]:		area	perimeter	width	height	convex_hull_area	longest_path
	1	25785.0	4788.927722	408	698	160303.0	4836
	2	35329.0	5155.881492	406	732	171239.5	5120
	3	24265.0	3272.229572	115	728	68062.0	4780
	4	408436.0	2820.000000	406	1006	407025.0	7430
	5	408438.0	2820.000000	406	1008	407025.0	7430
	6	119448.0	2982.315796	406	741	192712.5	5382
	7	408436.0	2820.000000	406	1006	407025.0	7430
	8	408436.0	2820.000000	406	1006	407025.0	7430
	9	30774.0	4905.923761	381	728	179952.5	5099
	10	27138.0	5079.267294	354	721	187842.0	5043
	11	408322.0	2821.899495	406	1008	407023.5	7428
	12	408436.0	2820.000000	406	1006	407025.0	7430
	13	408434.0	2819.414214	406	1008	407024.0	7428
	14	408436.0	2820.000000	406	1008	407025.0	7430
	15	408436.0	2820.000000	406	1006	407025.0	7430
	16	408436.0	2820.000000	406	1005	407025.0	7430
	17	101298.0	5085.143842	406	724	198039.5	5285
	18	408436.0	2820.000000	406	1006	407025.0	7430
	19	329472.0	8536.223561	406	1008	406982.5	7407



### Actual above-ground dry biomass

1	Notes	Block	Plant	# Ears	Ear Biomass	Mean Ear Bio	Plant and	Dry plant Seed (g)
2		ET	UN	9	18,212	2,023555556	21,286	10,669
3		ET	AN120	11	20,556	1,868727273	22,482	11,865
4		ET	ON	7	11,169	1,595571429	18,523	7,906
5	1	ET	NEO	8	14,676	1,8345	22,283	11,666
6		ET	ANN40	9	16,912	1,879111111	23,496	12,879



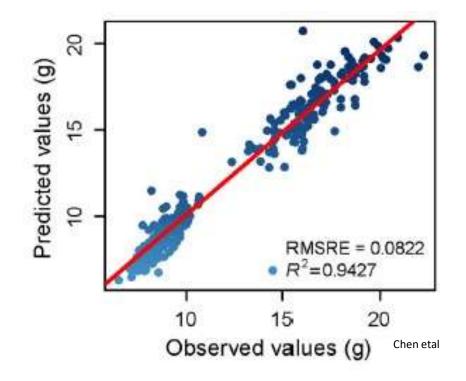
### **Prediction model**

- Train, validate and test
- MLR, PLSR, Deep neural networks for prediction
- Prediction model evaluation using R<sup>2</sup> and root-mean-square error (RMSE)



#### Predicted vs actual dry weight/biomass

 Image based predicted biomass vs manually measured biomass should be similar to this figure





### Conclusions

- Aa low-cost high-throughput image-based plant phenotyping system is developed using *Raspberry Pi* computer and *camera sensor* that captures plant image of shoots and characterizes plant traits to analyse plant growth dynamics.
- The study has demonstrated that image-based plant phenotyping is an efficient method to extract morphological features of plants (e.g., height, area, perimeter, etc).
- It has also shown that the area extracted from plant images can estimate plant growth rate.



## Thank you

